IN VITRO GROWTH RATE OF Kappaphycus alvarezii MICROPROPAGULE AND EMBRYO BY ENRICHMENT MEDIUM WITH SEAWEED EXTRACT

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ABSTRACT

The development of micropropagule and embryo of seaweed depend on nutrient and fertilizer used. Seaweed has been reported contain hormone regulators such as auxine, cytokinine, gibbereline, and various minerals applied in stimulating the growth ocra plant and wheat culture. The objectives of this study were to determine the potential of Kappaphycus alvarezii extract and its optimal concentration in accelerating of Kappaphycus alvarezii micropropagule and embryo growth. Micropropagule and embryo produced through callus induction were planted into PES 1/20 liquid medium supplemented with seaweed extract at the concentrations of 0 (control), 25, 50, 75, and 100 μL in 10 mL of medium. The results showed that medium enrichment with 50 μL of seaweed extract had the highest survival rate and growth of thallus. In addition, this concentration was also resulted in a good performance of K. alvarezii thallus with the lighter color. The advantage of this study for seaweed cultivation in Indonesia, among others, seaweed can be used as fertilizer, especially in the maintenance of seaweed seed, so that cultivation can be better develop.

KEYWORDS: callus induction, filamentous callus, micropropagule, embryo, Kappaphycus alvarezii

INTRODUCTION

Improvement the genetic quality of seaweed particularly Kappaphycus alvarezii has been demonstrated by using several methods such as hybrid izationin protoplasts or spores (Cheney & Emily, 1986) and genetic engineering carrageenan controller gene transfer or growth gene inserted in thallus, organ or embryo through mediation with agrobacterium (Hallman, 2007). The use of embryo as a medium for gene transfer is easier than the use of thallus or organ because it is a living cell resulted from callus induction and somatic embryo (Reddy et al., 2003). Gene transfer to the embryo can be implemented by using electroporator or gene pulser (Rajamuddin et al., 2010). Several promoters have been succesfully used in genetic engineering of seaweed such as cauliflower mosaic virus (CaMV), cytomegalovirus (CMV) and green fluorescent protein (GFP). Callus induction is one technique for finding out of seaweed embryo in vitro. The embryo development to be seedlings with callus induction through several phases and stages in which at the early stage is induction of explant grown to produce filaments. The callus induction method has been used by Reddy et al. (2003) on Kappaphycus alvarezii using a mixture of indole-3-acetic acid (IAA) and 6-benzylaminopurine (BAP) with several comparions in culture medium enriched with Provasoli’s enriched Seawater (PES) 1/20. Moreover, Suryati et al. (2006) have demonstrated callus induction and embryo by using Conway medium enriched with plant growth regulators (PGR) of a combination of IAA and kinetins the inductor. Embryos produced by using these media have been rudimentary, in which filaments produced have been not able to develop to be propagules and seedlings completely. Therefore, it is needed hormone regulators which can stimulate the development filaments to be intact and perfect seedlings.

Seaweed particularly Kappaphycus alvarezii has been reported contain PGR such as IAA, gyberelin GA3, kinetin, and zeatin, suspected to has the ability in accelerating of thallus forming on seaweed embro (Prasad et al., 2010). PGR isolated from seaweed extract were used in the multiplication of ocra plant and wheat culture (Zodape et al., 2008; 2009). In this study, the potential of K. alvarezii extract as growth regulator in accelerating the growth of K. alvarezii filaments and embryo was investigated.
MATERIALS AND METHODS

Seaweed Extract Preparation

Seaweed, Kappaphycus alvarezii used in this study was collected from Takalar Regency, South Sulawesi. Seaweeds were cleaned from dirt and epiphyte before washing with fresh water. The dirt was gently cleaned using a soft brush. Clean and salt free seaweeds were then ground to be slurry form and extracted with 90% methanol for 3 x 24 hours. Extract obtained was evaporated by using a rotary evaporator (Eyela type) to get dried methanol extract. The methanol extract was added with HCL 0.1 N to reach pH 2.5 and partitioned with ethyl acetate to yield ethyl acetate extract containing plant growth stimulators (PGR) of cytokinins and gibberelins derivative (Chapman & Chapman, 1980). Ethyl acetate extract resulted was then evaporated until reach 1 mL ethyl acetate extract equivalent to 100 g fresh weight.

Collection of Materials and Explant Preparation for Callus Induction

Kappaphycus alvarezii was collected from the farmer’s in Barru Regency. The sample was stored in the container covered by cloth moistened with sea water for further analysis. The analysis was conducted in the tissue culture laboratory of Research and Development Institute for Coastal Aquaculture (RDICA) Maros, South Sulawesi. The healthy and clean Kappaphycus alvarezii thallus were selected and cut with the size of 5 cm to be explants. The explants were then washed with sea water filtered with membrane filter and cultured for two weeks in sterile sea water enriched with PES 1/20 by using a shaker (IKA WERKE) at temperature of 22°C-25°C and illuminated with fluorescent lamps at 1,500 lux of light intensity with a 12:12 hours light and dark cycle. During two weeks of rearing time period, 10 mg/L GeO₂ was mixed into culture medium to remove diatoms. The cultured explants for callus induction were sterilized by using surface sterilization method as described by Polne & Gibor (1984) and Huang & Fujita (1997) with some modification. The explants were gently cleaned using a soft brush under a stereoscopic microscope. The explants sterilization was carried out by washing the explants using 0.5% liquid detergent in sterile sea water for 10 minutes. The explants were then treated with 2% betadine in the sterile sea water for three minutes and continued to be sterilized with 3% broad spectrum antibiotic mixture in PES culture medium for two days. To confirm the sterilization, the culture medium was inoculated on agar medium and incubated.

Callus Induction

The sterile explants were washed with sterile sea water and cut into pieces with 4-5 mm length. Each explant was wiped gently onsterile filter papers to remove moisture and any mucilaginous substances exuded from the cut ends. Dried explants were planted on 20 mL of 0.8% (w/v) Bacto-Agar-solidified PES 1/20 medium enriched with seaweed extract at the concentrations of 0 (control), 25, 50, 75, and 100 μL with the density of 15 explants/petri dish and cultured in the glass cupboard at the temperature of 20°C using a neon lamp at 1,500 lux with photoperiod of dark and light (12 hours). Each treatment was applied with 10 replications. After 30 days of callus induction, the transparent filaments and brown filaments formed on the explant were counted their number and growth. Subsequently, all callus out growth were excised from the explants and subcultured separately on 20 mL of 0.5% (w/v) Bacto-Agar-solidified PES 1/20 medium enriched with seaweed extract at the concentrations of 0, 25, 50, 75, and 100 μL. After 30 days of rearing time periods, all callus separately cultured resulted in micropropagules. During the rearing of callus into micropropagules, the development of filamentous callus was observed under a microscope (Bausch & Lomb).

Embryo Differentiation

Embryo differentiation was carried out by subculturing of micropropagules produced into PES 1/20 liquid medium supplemented with seaweed extract containing plant growth regulators (PGR) from sitokinin and giberelin derivative at the concentration of 0 (control), 25, 50, 75, and 100 μL. The cultures were shaken with a shaker for 60 days at room temperature of 22°C-25°C and illuminated with fluorescent lamps at 1,500 lux of light intensity with a 12 hours light and dark cycle. All treatments were carried out in 10 replicates. The micropropagules were reared for 60 days or until yielded thallus with medium change interval of two weeks. The data were collected during the observation such as the condition, mortality, and growth of thallus. The data were analyzed using ANOVA statistical method.

RESULTS AND DISCUSSIONS

To stimulate the differentiation of embryonic seaweed K. alvarezii to the talus, in addition used of commercial PGR, in this study also used K. alvarezii seaweed extract to stimulate the growth and differentiation of the talus filament to be perfect. Prasad et al. (2010) have demonstrated on detection the presence of a class of plant growth regulator auxin and
cytokinin in the seaweed extract. Zodape et al. (2008; 2009) have taken advantage of this seaweed extract on okra plants and cultivation of wheat. In addition, some researcher have conducted assessments of fertilizer derived from seaweed Ascophyllum nodosum, among others, which can be used as fertilizer in cultivation of seaweed K. alvarezii (Loureiro et al., 2010) and to test the animal manure livestock, as well as fertilizer in higher plants (Khan et al., 2011).

At embryonic differentiation of seaweed, seaweed extract was used at concentrations ranging from 0-100 μL containing 1 mL of extract equivalent to 100 grams of fresh weight. The results show that the growth and survival of embryos seaweed K. alvarezii on 1/20 PES culture medium enriched with seaweed extracts 50 μL in 10 mL of culture medium had the highest survival of about 82%, while in the control (no addition of extract) had the lowest survival and tend to blanch which eventually die (Figure 1).

Figure 1 shows that the highest survival rate (82%) of K. alvarezii embryo was obtain from the concentration of 50 μL in 10 mL culture medium (1 mL extract equivalent with 100 g fresh weight of seaweed), followed by 75, 100, 25, and 0 μL with the survival rate of 60%, 58%, 42%, and 20%, respectively. Compared to the control, (without seaweed extract addition), survival rate exhibited by K. alvarezii embryo planted into PES 1/20 liquid medium enriched with seaweed extract at all concentrations was indicated a similar pattern in which the growth rate drastically increased after a week of rearing time periods and continue to gradually increased until eight weeks of rearing time period. The statistical analysis showed that treatment 50 μL/L provides the highest value is 82.30% significantly different from the control (not given the extract) with a value of 21%; and 25 μL/L with a value of 42%, but not significantly different from the treatment 75 μL/L with a
The enrichment of PES medium with 50 μL of seaweed extract was also indicated a good response on the filament sand embryos growth in which the phase of filament into callus, callus into thallus grew faster with lighter colour (Figure 3).

The filament growth was also influenced by culture media and nutrient content in the culture media. Medium enrichment with seaweed extract in this study was able to stimulate the filament growth due to the presence of hormones and nutrients in seaweed extract as reported by Prasad et al. (2010). K. alvarezii produced nutrients and hormones which have a function in the growth and survival rate of seaweed. Seaweed extract was used as growth regulators for ocra plant and wheat culture. Nevertheless, seaweed extract used in stimulating the growth of K. alvarezii was reported for the first time in this study.

CONCLUSIONS

*Kappaphycus alvarezii* extract has the potential to be developed as growth hormone to stimulate the growth of filament into callus and callus into thallus. PES 1/20 culture medium enriched with seaweed extract at the concentration of 50 μL/10 mL culture medium was indicate the highest survival rate and growth of *Kappaphycus alvarezii* filaments and embryos and also produced filaments with a good and lighter performance. It is suggested to isolate and identify hormone regulators used from *Kappaphycus alvarezii*.

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